



High Performance Computing for genomic applications Using genomic software on Euler

Scientific IT Services
Michał Okoniewski

Bioinformatic modules on EULER

- module load
- module load gcd
- module avail
- module purge
- env2lmod, lmod2env
- https://scicomp.ethz.ch/wiki/New_SPACK_software_stack_on_Euler



michalo — michalo@eu-login-01:~ — tmux — 95x21

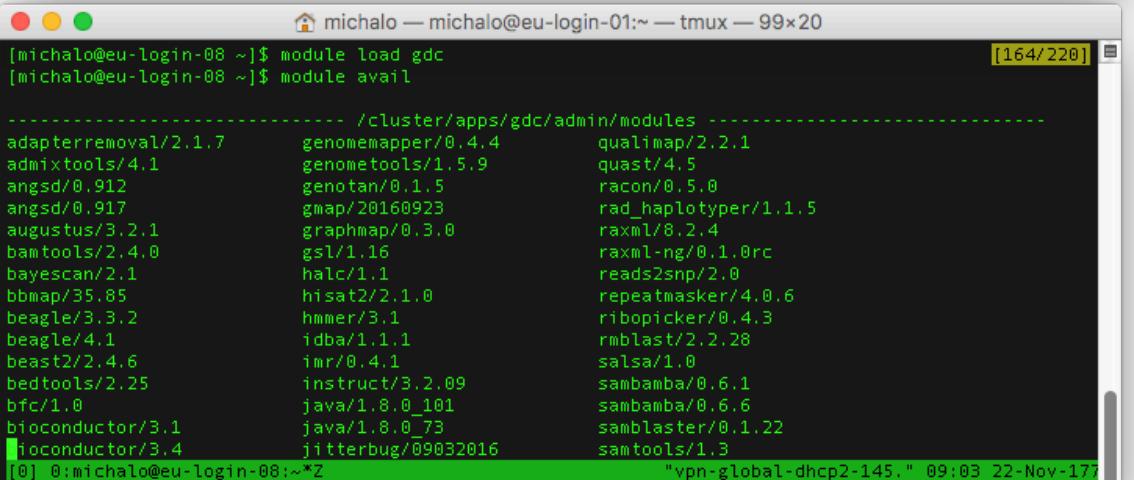
Eidgenössische Technische Hochschule Zuerich
Swiss Federal Institute of Technology Zurich

E U L E R C L U S T E R

<https://scicomp.ethz.ch>
<http://tinyurl.com/cluster-support>
cluster-support@id.ethz.ch

```
[michalo@eu-login-08 ~]$ module load bioconductor/3.4
bioconductor/3.0(default:3) bioconductor/3.4
[michalo@eu-login-08 ~]$ module load bioconductor/3.4
Using OpenBLAS build of bioconductor R-3.4
[michalo@eu-login-08 ~]$
```

[0] 0:michalo@eu-login-08:~*Z "vpn-global-dhcp2-145." 09:01 22-Nov-17



michalo — michalo@eu-login-01:~ — tmux — 99x20 [164/220]

```
[michalo@eu-login-08 ~]$ module load gcd
[michalo@eu-login-08 ~]$ module avail
```

----- /cluster/apps/gdc/admin/modules -----

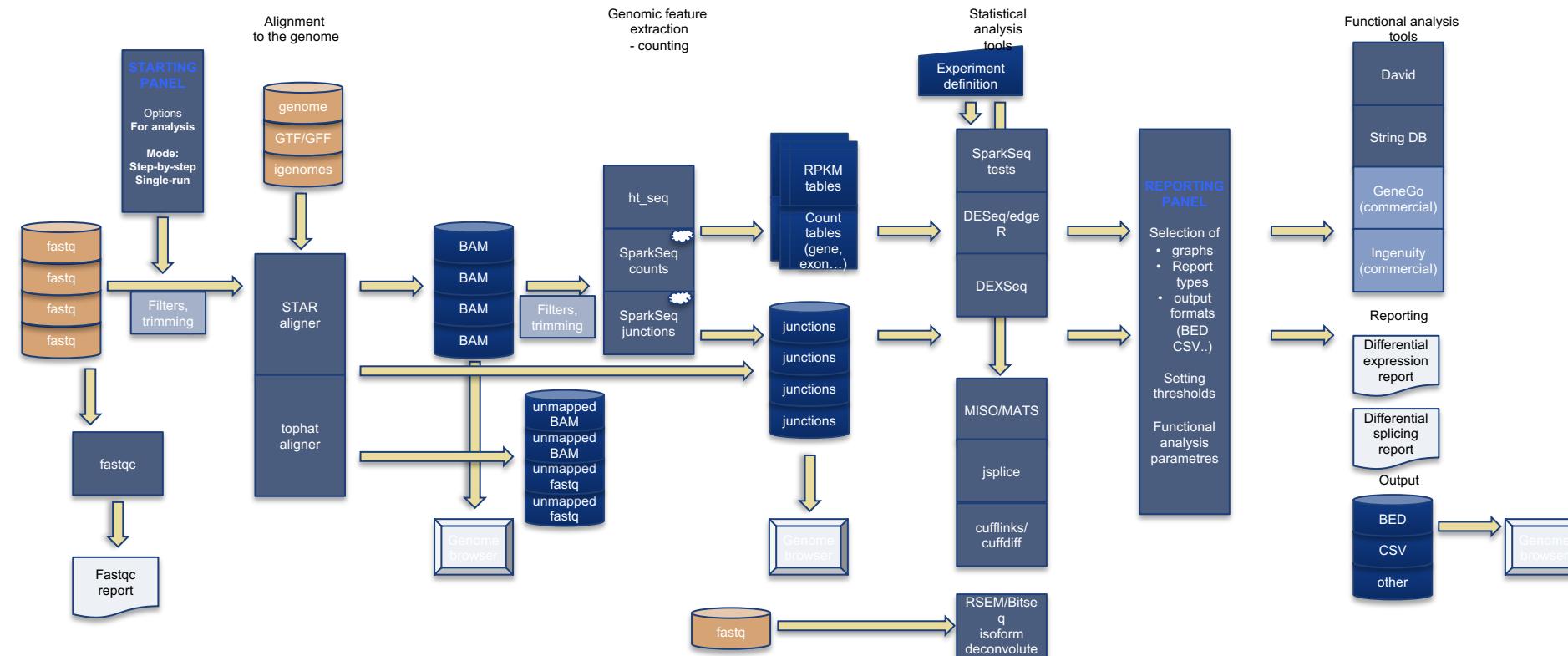
adapterremoval/2.1.7	genomemapper/0.4.4	qualimap/2.2.1
admixtools/4.1	genometools/1.5.9	quast/4.5
angsd/0.912	genotan/0.1.5	racon/0.5.0
angsd/0.917	gmap/20160923	rad_haplotype/1.1.5
augustus/3.2.1	graphmap/0.3.0	raxml/8.2.4
bamtools/2.4.0	gsl/1.16	raxml-ng/0.1.0rc
bayescan/2.1	halc/1.1	reads2snp/2.0
bbmap/35.85	hisat2/2.1.0	repeatmasker/4.0.6
beagle/3.3.2	hmmer/3.1	ribopicker/0.4.3
beagle/4.1	idba/1.1.1	rmblast/2.2.28
beast2/2.4.6	imr/0.4.1	salsa/1.0
bedtools/2.25	instruct/3.2.09	sambamba/0.6.1
bfc/1.0	java/1.8.0_101	sambamba/0.6.6
bioconductor/3.1	java/1.8.0_73	samblaster/0.1.22
bioconductor/3.4	jitterbug/09032016	samtools/1.3

```
[0] 0:michalo@eu-login-08:~*Z "vpn-global-dhcp2-145." 09:03 22-Nov-17
```

Bioinformatic software jungle - categories

- Trimming tools: trimmomatic, cutadapt
- Aligners:
 - General purpose: bwa, bowtie, SHRiMP
 - RNA aligners: STAR, tophat, subjunc, hisat2
 - Transcriptome aligners: kallisto, salmon, sailfish, RSEM
 - Other aligners: Blast, Blat, VMATCH
- De-novo assemblers: trinity, velvet, spades
- Feature extraction, counting: HTSeq, featureCount
 - Transcript discovery: cufflinks,
- Specialized tools: MISO, blast2go,...
- Utilities and conversion tools: samtools, bcftools, bedtools, picard tools

Bioinformatic software jungle (as seen back in 2015)



Trimming tools

- trimmomatic, cutadapt
- Trimming:
 - Removing adapters
 - By quality: eg sliding window
 - Fixed position, eg several base pairs at each end
- Typically: fastq on input, fastq on output
 - it gets more complex with paired reads

Trimming tools

```
module load java  
module load trimmomatic
```

```
trimmomatic SE {input} {output} ILLUMINA_CLIP:adapters.fa:2:30:10  
LEADING:3 TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36
```

```
>TruSeqR2_nextera  
CTGTCTCTTATACACATCTCGAGCCCACGAGACTAAGGCGAATCTGTATGCCCTCTGCTTGAAAA  
>nextera_R2_right_side_adapter  
CTGTCTCTTATACACATCTGACGCTGCCGACGAGCGATCTAGTAGATCTGGTGGTCGCCGTATCATTAAAA  
>I5_Nextera_Transposase_1  
CTGTCTCTTATACACATCTGACGCTGCCGACGA  
>I7_Nextera_Transposase_1  
CTGTCTCTTATACACATCTCGAGCCCACGAGAC  
>I5_Nextera_Transposase_2  
CTGTCTCTTATACACATCTGATGGCGCGAGGGAGGC  
>I7_Nextera_Transposase_2  
CTGTCTCTTATACACATCTGAGCAGGGCTGGCAAGGC  
>I5_Primer_Nextera_XT_and_Nextera_Enrichment_[N/S/E]501  
GACGCTGCCGACGAGCGATCTAGTAGATCTGGTGGTCGCCGTATCATT  
>I5_Primer_Nextera_XT_and_Nextera_Enrichment_[N/S/E]502_and_I5_Primer_Nextera_XT_Index_Kit_v2_S502  
GACGCTGCCGACGAATAGAGAGGTGAGATCTGGTGGTCGCCGTATCATT  
>I5_Primer_Nextera_XT_and_Nextera_Enrichment_[N/S/E]503_and_I5_Primer_Nextera_XT_Index_Kit_v2_S503  
GACGCTGCCGACGAAGAGGATAGTAGATCTGGTGGTCGCCGTATCATT  
>I5_Primer_Nextera_XT_and_Nextera_Enrichment_[N/S/E]504  
GACGCTGCCGACGATCTACTCTGTAGATCTGGTGGTCGCCGTATCATT  
>I5_Primer_Nextera_XT_and_Nextera_Enrichment_[N/S/E]505_and_I5_Primer_Nextera_XT_Index_Kit_v2_S505  
GACGCTGCCGACGACTCTTACGTGAGATCTGGTGGTCGCCGTATCATT  
>I5_Primer_Nextera_XT_and_Nextera_Enrichment_[N/S/E]506_and_I5_Primer_Nextera_XT_Index_Kit_v2_S506  
GACGCTGCCGACGATATCAGTAGATCTGGTGGTCGCCGTATCATT  
>I5_Primer_Nextera_XT_and_Nextera_Enrichment_[N/S/E]507_and_I5_Primer_Nextera_XT_Index_Kit_v2_S507  
GACGCTGCCGACGATACTCCTTGTAGATCTGGTGGTCGCCGTATCATT
```



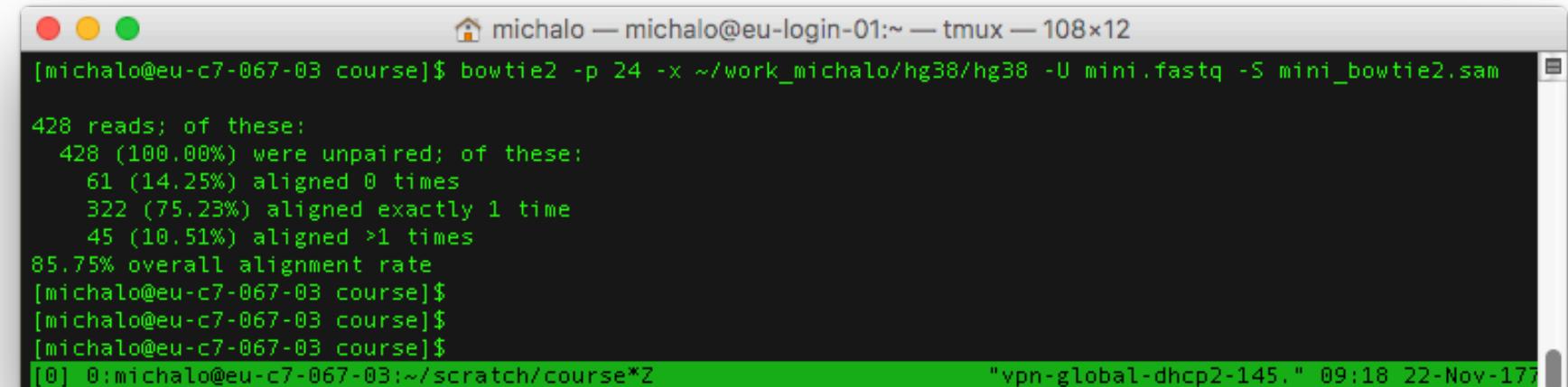
Bowtie, bowtie2

- Building genome index

Read: GACTGGGCGATCTCGACTTCG
||||| ||||| ||||| |||
Reference: GACTG--CGATCTCGACATCG

```
bowtie2-build --threads 24
/cluster/home/michalo/work_michalo/hg38/Homo_sapiens.GRCh38.dna.primary_assembly.fa
/cluster/scratch/michalo/hg38/hg38
```

- Alignment:



A screenshot of a terminal window titled "michalo — michalo@eu-login-01:~ — tmux — 108x12". The window displays the output of a "bowtie2" command. The output shows the following statistics:

```
[michalo@eu-c7-067-03 course]$ bowtie2 -p 24 -x ~/work_michalo/hg38/hg38 -U mini.fastq -S mini_bowtie2.sam
428 reads; of these:
  428 (100.00%) were unpaired; of these:
    61 (14.25%) aligned 0 times
    322 (75.23%) aligned exactly 1 time
    45 (10.51%) aligned >1 times
  85.75% overall alignment rate
[michalo@eu-c7-067-03 course]$
[michalo@eu-c7-067-03 course]$
[michalo@eu-c7-067-03 course]$
[0] 0:michalo@eu-c7-067-03:~/scratch/course*2          "vpn-global-dhcp2-145." 09:18 22-Nov-17
```

Tophat

- Classic splice-aware aligner
- Uses bowtie2 as engine, so also bowtie2 index

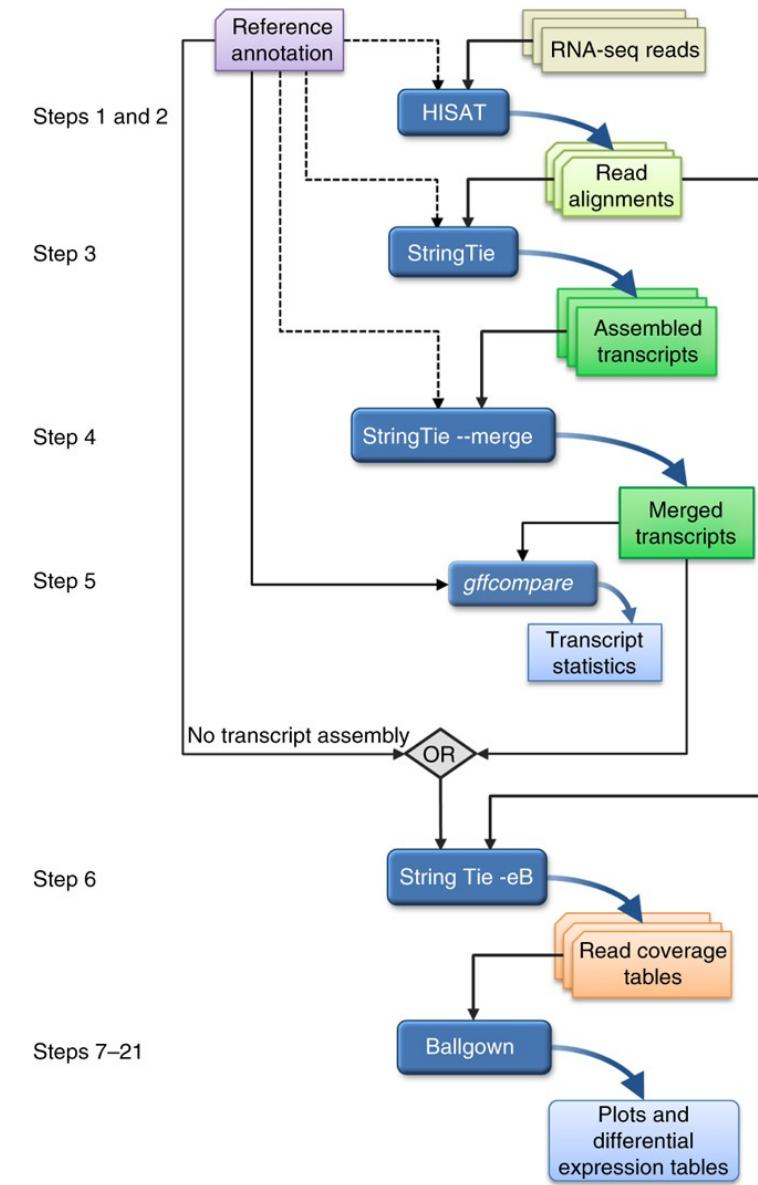
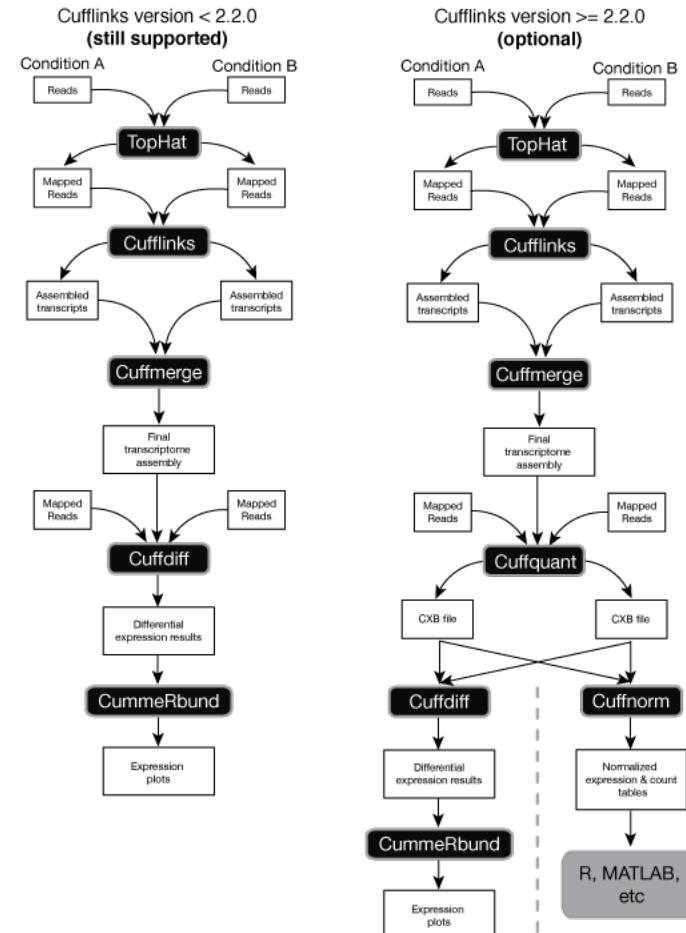
```
tophat -p 24 -o tophat_out --library-type fr-firststrand ~/work_michalo/hg38/hg38 mini.fastq.gz
```

- Manual: <http://ccb.jhu.edu/software/tophat/index.shtml>

Splice aware aligners for RNA-seq, memory needs

	Index type	Index size=memory needs for hg38	Computing node
Tophat, 2011	bowtie index in a file	in a file	any
STAR, 2014	In memory	ca 40GB	EULER normal (65G)
Subjunc, 2014	In memory	ca 40GB	EULER normal (65G)
Hisat2, 2016	In memory	ca 7GB	good laptop (>=8G RAM)

Tuxedo suite and “new tuxedo”



“New tuxedo suite”



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Published: 11 August 2016

Transcript-level expression analysis of RNA-seq experiments with HISAT, StringTie and Ballgown

Mihaela Pertea, Daehwan Kim, Geo M Pertea, Jeffrey T Leek & Steven L Salzberg 

Nature Protocols 11, 1650–1667(2016) | Cite this article

43k Accesses | 1133 Citations | 87 Altmetric | Metrics

Cufflinks and stringtie

- Transcript discovery tools
- Uses coverage and junctions from a BAM file

```
cufflinks mini_star.sorted.bam
```

- Other
 - cuffmerge, cuffdiff, cuffquant, cuffnorm, CummeRbund

Cufflinks and stringtie

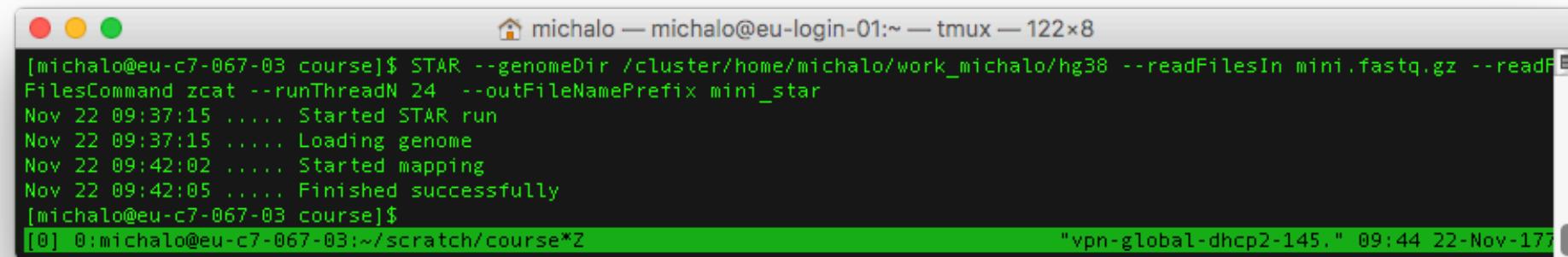
- Produces GTF

```
michalo — michalo@eu-login-01:~— tmux — 265x41
17 Cufflinks transcript 41801952 41802294 1000 . .
17 Cufflinks exon 41801952 41802294 1000 . .
17 Cufflinks transcript 41802365 41803426 337 . .
17 Cufflinks exon 41802365 41802981 337 . .
17 Cufflinks exon 41803287 41803426 337 . .
17 Cufflinks transcript 41802365 41803426 1000 . .
17 Cufflinks exon 41802365 41802979 1000 . .
17 Cufflinks exon 41803287 41803426 1000 . .
17 Cufflinks transcript 41806622 41809842 1000 . .
17 Cufflinks exon 41806622 41806879 1000 . .
17 Cufflinks exon 41807859 41808084 1000 . .
17 Cufflinks exon 41809706 41809842 1000 . .
17 Cufflinks transcript 41810873 41811019 1000 . .
17 Cufflinks exon 41810873 41811019 1000 . .
17 Cufflinks transcript 41811134 41811385 1000 . .
17 Cufflinks exon 41811134 41811385 1000 . .
17 Cufflinks transcript 41811684 41811948 1000 . .
17 Cufflinks exon 41811684 41811948 1000 . .
17 Cufflinks transcript 41825205 41835234 1000 . .
17 Cufflinks exon 41825205 41825657 1000 . .
17 Cufflinks exon 41827426 41827626 1000 . .
17 Cufflinks exon 41828798 41828952 1000 . .
17 Cufflinks exon 41830801 41830890 1000 . .
17 Cufflinks exon 41832392 41832477 1000 . .
17 Cufflinks exon 41835070 41835116 1000 . .
17 Cufflinks exon 41835203 41835234 1000 . .
17 Cufflinks transcript 41825205 41836162 306 . .
17 Cufflinks exon 41825205 41825657 306 . .
17 Cufflinks exon 41827426 41827626 306 . .
17 Cufflinks exon 41828798 41828952 306 . .
17 Cufflinks exon 41830801 41830890 306 . .
17 Cufflinks exon 41832392 41832477 306 . .
17 Cufflinks exon 41835070 41835272 306 . .
17 Cufflinks exon 41835859 41836162 306 . .
. . . . . .
gene_id "CUFF.1"; transcript_id "CUFF.1.1"; FPKM "385640.0529891439"; frac "1.000000"; conf_lo "224014.123428"; conf_hi "547265.982551"; cov "8.014439";
gene_id "CUFF.1"; transcript_id "CUFF.1.1"; exon_number "1"; FPKM "385640.0529891439"; frac "1.000000"; conf_lo "224014.123428"; conf_hi "547265.982551"; cov "8.014439";
gene_id "CUFF.2"; transcript_id "CUFF.2.1"; FPKM "61688.3599707013"; frac "0.252522"; conf_lo "20863.162216"; conf_hi "111513.557726"; cov "1.382272";
gene_id "CUFF.2"; transcript_id "CUFF.2.1"; exon_number "1"; FPKM "61688.3599707013"; frac "0.252522"; conf_lo "20863.162216"; conf_hi "111513.557726"; cov "1.382272";
gene_id "CUFF.2"; transcript_id "CUFF.2.1"; exon_number "2"; FPKM "61688.3599707013"; frac "0.252522"; conf_lo "20863.162216"; conf_hi "111513.557726"; cov "1.382272";
gene_id "CUFF.2"; transcript_id "CUFF.2.2"; FPKM "195920.7516271057"; frac "0.747478"; conf_lo "129358.547713"; conf_hi "262482.955541"; cov "4.091593";
gene_id "CUFF.2"; transcript_id "CUFF.2.2"; exon_number "2"; FPKM "195920.7516271057"; frac "0.747478"; conf_lo "129358.547713"; conf_hi "262482.955541"; cov "4.091593";
gene_id "CUFF.3"; transcript_id "CUFF.3.1"; FPKM "399883.8092033406"; frac "1.000000"; conf_lo "222379.790173"; conf_hi "559587.828234"; cov "8.190210";
gene_id "CUFF.3"; transcript_id "CUFF.3.1"; exon_number "1"; FPKM "399883.8092033406"; frac "1.000000"; conf_lo "222379.790173"; conf_hi "559587.828234"; cov "8.190210";
gene_id "CUFF.3"; transcript_id "CUFF.3.1"; exon_number "2"; FPKM "399883.8092033406"; frac "1.000000"; conf_lo "222379.790173"; conf_hi "559587.828234"; cov "8.190210";
gene_id "CUFF.3"; transcript_id "CUFF.3.1"; exon_number "3"; FPKM "399883.8092033406"; frac "1.000000"; conf_lo "222379.790173"; conf_hi "559587.828234"; cov "8.190210";
gene_id "CUFF.4"; transcript_id "CUFF.4.1"; FPKM "2721172.9130021906"; frac "1.000000"; conf_lo "1080243.905346"; conf_hi "4362101.920658"; cov "56.977279";
gene_id "CUFF.4"; transcript_id "CUFF.4.1"; exon_number "1"; FPKM "2721172.9130021906"; frac "1.000000"; conf_lo "1080243.905346"; conf_hi "4362101.920658"; cov "56.977279";
gene_id "CUFF.5"; transcript_id "CUFF.5.1"; FPKM "386460.4071371864"; frac "1.000000"; conf_lo "136927.788094"; conf_hi "635993.026180"; cov "8.059207";
gene_id "CUFF.5"; transcript_id "CUFF.5.1"; exon_number "1"; FPKM "386460.4071371864"; frac "1.000000"; conf_lo "136927.788094"; conf_hi "635993.026180"; cov "8.059207";
gene_id "CUFF.6"; transcript_id "CUFF.6.1"; FPKM "365523.9830166674"; frac "1.000000"; conf_lo "145104.727827"; conf_hi "585943.238206"; cov "7.632440";
gene_id "CUFF.6"; transcript_id "CUFF.6.1"; exon_number "1"; FPKM "365523.9830166674"; frac "1.000000"; conf_lo "145104.727827"; conf_hi "585943.238206"; cov "7.632440";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "1"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "2"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "3"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "4"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "5"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "6"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.1"; exon_number "7"; FPKM "386421.1203223147"; frac "0.686190"; conf_lo "311028.834987"; conf_hi "461813.405657"; cov "8.087935";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; exon_number "1"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; exon_number "2"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; exon_number "3"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; exon_number "4"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; exon_number "5"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; exon_number "6"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
gene_id "CUFF.7"; transcript_id "CUFF.7.2"; exon_number "7"; FPKM "118402.6643432135"; frac "0.313810"; conf_lo "81225.981370"; conf_hi "155579.347316"; cov "2.478211";
(END)
[8] 0:michalo@eu-c7-067-03:~/scratch/course*2
 vpn-global-dhcp2-145. 10:06 22-Nov-17
```



STAR

- Splice aware aligner, loading index into memory
- Results similar to tophat, but faster
- --genomeLoad LoadAndKeep
- With specific options, can produce BAM and do the counting too



A screenshot of a Mac OS X terminal window titled "michalo — michalo@eu-login-01:~ — tmux — 122x8". The window shows the command [michalo@eu-c7-067-03 course]\$ STAR --genomeDir /cluster/home/michalo/work_michalo/hg38 --readFilesIn mini.fastq.gz --readFilesCommand zcat --runThreadN 24 --outFileNamePrefix mini_star being run. The output shows the process starting at Nov 22 09:37:15, loading the genome, starting mapping, and finishing successfully at Nov 22 09:42:05. The terminal prompt [michalo@eu-c7-067-03 course]\$ is visible at the bottom, along with the session identifier [0] and the date "22-Nov-17".

- <https://github.com/alexdobin/STAR/blob/master/doc/STARmanual.pdf>

STAR statistics

```
michalo — michalo@eu-login-01:~ — tmux — 82x33
      Started job on | Nov 22 09:37:15
      Started mapping on | Nov 22 09:42:02
      Finished on | Nov 22 09:42:05
      Mapping speed, Million of reads per hour | 0.51

      Number of input reads | 428
      Average input read length | 49
      UNIQUE READS:
      Uniquely mapped reads number | 419
      Uniquely mapped reads % | 97.90%
      Average mapped length | 48.74
      Number of splices: Total | 47
      Number of splices: Annotated (sjdb) | 0
      Number of splices: GT/AG | 47
      Number of splices: GC/AG | 0
      Number of splices: AT/AC | 0
      Number of splices: Non-canonical | 0
      Mismatch rate per base, % | 0.41%
      Deletion rate per base | 0.00%
      Deletion average length | 0.00
      Insertion rate per base | 0.00%
      Insertion average length | 0.00
      MULTI-MAPPING READS:
      Number of reads mapped to multiple loci | 9
      % of reads mapped to multiple loci | 2.10%
      Number of reads mapped to too many loci | 0
      % of reads mapped to too many loci | 0.00%
      UNMAPPED READS:
      % of reads unmapped: too many mismatches | 0.00%
      % of reads unmapped: too short | 0.00%
      % of reads unmapped: other | 0.00%
(END)
[0] <chalo@eu-c7-067-03:~/scratch/course*2 "vpn-global-dhcp2-145." 09:46 22-Nov-177
```

subread

- Includes subjunc similar to STAR and featureCounts
- Building index

```
subread-buildindex -o /cluster/home/michalo/work_michalo/hg38/subread_index/hg38  
/cluster/home/michalo/work_michalo/hg38/Homo_sapiens.GRCh38.dna.primary_assembly.fa
```

- Alignment

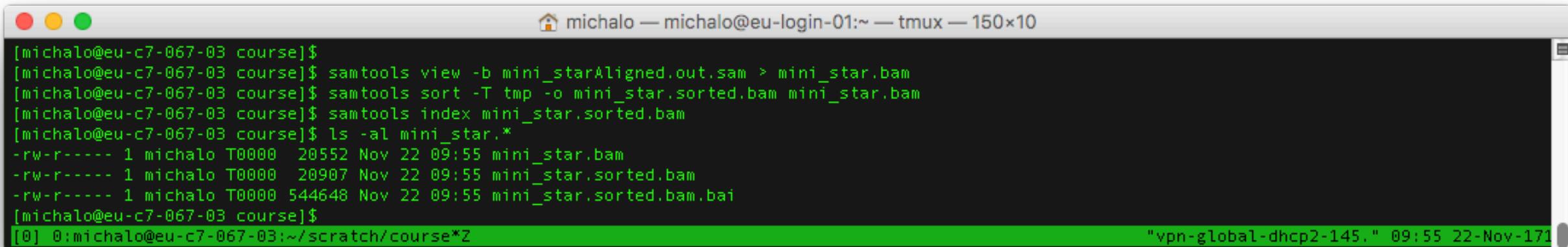
```
subread -T 24 -i /cluster/home/michalo/work_michalo/hg38/subread_index/hg38 -r mini.fastq -o  
mapped_reads_subjunc/mini.bam
```

```
subjunc -T 24 -i /cluster/home/michalo/work_michalo/hg38/subread_index/hg38 -r mini.fastq -o  
mapped_reads_subjunc/mini.bam
```

- <http://bioinf.wehi.edu.au/subread-package/SubreadUsersGuide.pdf>

samtools

- General purpose tool for conversion of BAM ⇔ SAM
- Many other operations: pileup...
- See: <http://www.htslib.org/doc/samtools.html>



```
michalo — michalo@eu-login-01:~ — tmux — 150x10
[michalo@eu-c7-067-03 course]$
[michalo@eu-c7-067-03 course]$ samtools view -b mini_starAligned.out.sam > mini_star.bam
[michalo@eu-c7-067-03 course]$ samtools sort -T tmp -o mini_star.sorted.bam mini_star.bam
[michalo@eu-c7-067-03 course]$ samtools index mini_star.sorted.bam
[michalo@eu-c7-067-03 course]$ ls -al mini_star.*
-rw-r----- 1 michalo 10000 20552 Nov 22 09:55 mini_star.bam
-rw-r----- 1 michalo 10000 20907 Nov 22 09:55 mini_star.sorted.bam
-rw-r----- 1 michalo 10000 544648 Nov 22 09:55 mini_star.sorted.bam.bai
[michalo@eu-c7-067-03 course]$
[0] 0:michalo@eu-c7-067-03:~/scratch/course*2
"vpn-global-dhcp2-145." 09:55 22-Nov-171
```

featureCounts

- Fast and flexible counting in genomic features

```
featureCounts -M -s 2 -T 24 -t gene -g gene_id -a  
/cluster/home/michalo/work_michalo/hg38/Homo_sapiens.GRCh38.86.chr.gtf -o mini.cnt  
mini_star.sorted.bam
```

featureCounts

```
michalo — michalo@eu-login-01:~ — tmux — 111x49
[michalo@eu-c7-067-03 course]$ featureCounts -M -s 2 -T 24 -t gene -g gene_id -a /cluster/home/michalo/work_michalo/hg38/Homo_sapiens.GRCh38.86.chr.gtf -o mini.cnt mini_star.sorted.bam
===== SUBREAD =====
v1.5.0

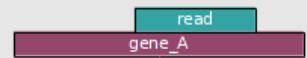
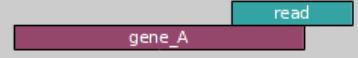
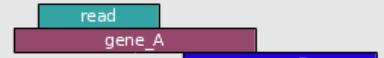
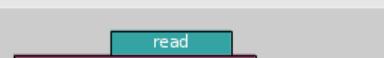
//----- featureCounts setting -----
|| Input files : 1 BAM file
||           5 mini_star.sorted.bam
|| Output file : mini.cnt
|| Annotations : /cluster/home/michalo/work_michalo/hg38/Homo ...
|| Threads : 16
|| Level : meta-feature level
|| Paired-end : no
|| Strand specific : inverted
|| Multi-mapping reads : counted (as integer)
|| Multi-overlapping reads : not counted
|| Read orientations : fr
\\----- http://subread.sourceforge.net/ -----//

//----- Running -----
|| Load annotation file /cluster/home/michalo/work_michalo/hg38/Homo_sapi ...
|| Features : 57992
|| Meta-features : 57992
|| Chromosomes/contigs : 25
|| Process BAM file mini_star.sorted.bam...
|| Single-end reads are included.
|| Assign reads to features...
|| Total reads : 450
|| Successfully assigned reads : 401 (89.1%)
|| Running time : 0.00 minutes
|| Read assignment finished.
\\----- http://subread.sourceforge.net/ -----//

[michalo@eu-c7-067-03 course]$
```

```
michalo — michalo@eu-login-01:~ — tmux — 67x19
Status mini_star.sorted.bam
Assigned 401
Unassigned_Ambiguity 1
Unassigned_MultiMapping 0
Unassigned_NoFeatures 48
Unassigned_Unmapped 0
Unassigned_MappingQuality 0
Unassigned_FragmentLength 0
Unassigned_Chimera 0
Unassigned_Secondary 0
Unassigned_Nonjunction 0
Unassigned_Duplicate 0
^
^
^
^
^
^
(END)
[0] <-03:~/scratch/course*Z "vpn-global-dhcp2-145." 10:00 22-Nov-17
```

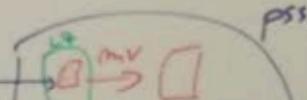
Counting reads in genes is non-trivial!

	union	intersection _strict	intersection _nonempty
 (gene_A)	gene_A	gene_A	gene_A
 (gene_A)	gene_A	no_feature	gene_A
 (gene_A, gene_A)	gene_A	no_feature	gene_A
 (gene_A, gene_A)	gene_A	gene_A	gene_A
 (gene_A, gene_B)	gene_A	gene_A	gene_A
 (gene_A, gene_B)	ambiguous (both genes with --nonunique all)	gene_A	gene_A
 (gene_A, gene_B)	ambiguous (both genes with --nonunique all)		
 (gene_A, gene_B)	alignment_not_unique (both genes with --nonunique all)		

htSeq counting modes
by Simon Anders

Tuning featureCounts

- Defaults counting features can be found in current STAR
- Still, for many cases a more careful counting is needed
- Gene or exon level counting
- Options

defaut (unique)	- M (with MatMap)	- M - O (with overlap) FLAG: ✓ NH:i: ✓	geneCount (STAR) = HTSeq-unique FLAG: O + 16 NH:i: 1 or MAPQ > 255
	pss5		

SRA tools

The command line download of public datasets from Short Read Archive

<http://ncbi.github.io/sra-tools/>

fastq-dump

fastq-dump --split-files ERR2811092

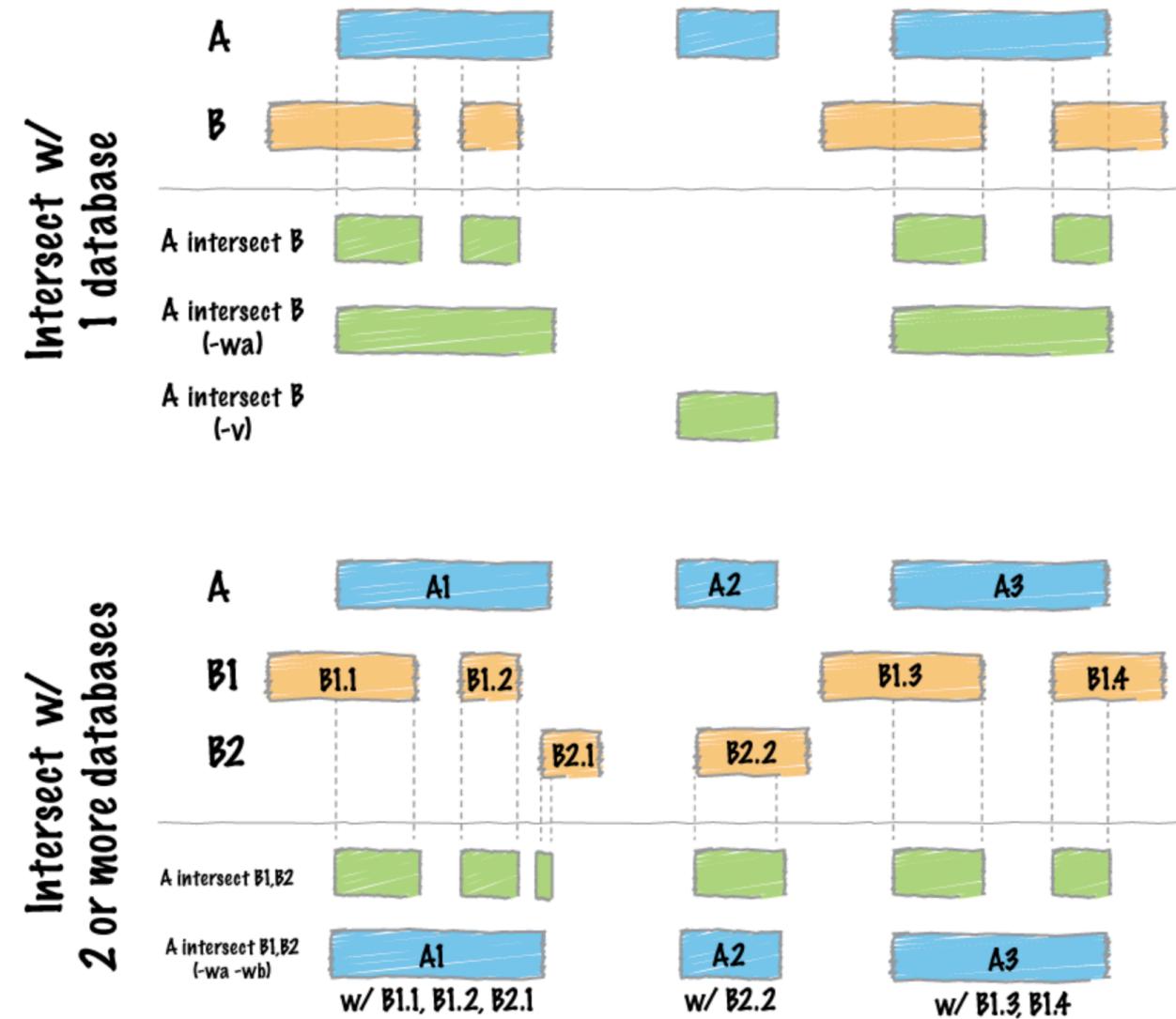
(from v2.9) fasterq-dump --split-files ERR2811092 -e 4

Bedtools

- “swiss army knife” for BED, GFF, SAM/BAM, VCF
- Conversion
- Annotation
- Getting sequences
- Genomic ranges
-



Bedtools - intersect



GATK

- GATK is a genomic toolbox for various operations related mainly to genomic variants calling
- Operations include producing a variant file *.vcf from an alignment file *.bam

```
module load gcc/4.8.2 gdc java/1.8.0_73
gatk/3.5java -jar GenomeAnalysisTK.jar -T UnifiedGenotyper -R
ref/human_g1k_b37_20.fasta -I bams/exp_design/NA12878_wgs_20.bam -o
sandbox/NA12878_wgs_20_UG_calls.vcf -glm BOTH -L 20:10,000,000-10,200,000
```

<https://software.broadinstitute.org/gatk/documentation/tooldocs/current/>

<https://software.broadinstitute.org/gatk/documentation/topic?name=tutorials>

<http://gatkforums.broadinstitute.org/gatk/discussion/7869/howto-discover-variants-with-gatk-a-gatk-workshop-tutorial>

Bioinformatics software stack on EULER

- [https://scicomp.ethz.ch/wiki/GDC software stack](https://scicomp.ethz.ch/wiki/GDC_software_stack)
- Commands to call modules for a specific tool
- List compiled in collaboration with Genomic Diversity Center from D-USYS

Resources needed by bioinformatic tools on EULER

		cores	memory node	time/queue
Trimming	Trimmomatic, cutadapt	1		24h
Aligners - old	bowtie, bwa, SHRiMP, tophat	many: 24		4h
Aligners - RNAseq	STAR, subread	many: 24	65G	4h
Aligners - optimized	hisat2	many: 24		4h
Conversion tools	Samtools, bcftools	1		4h
Counting	featureCount	1		4h
De-novo assembly	Spades, velvet	many	256GB-1T	24h or more
Variant calling	GATK, pileup	many	256GB	24h
...				

Thank you!

Using genomic software on Euler

